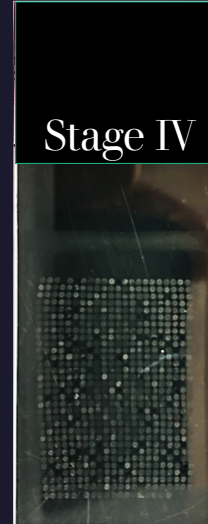
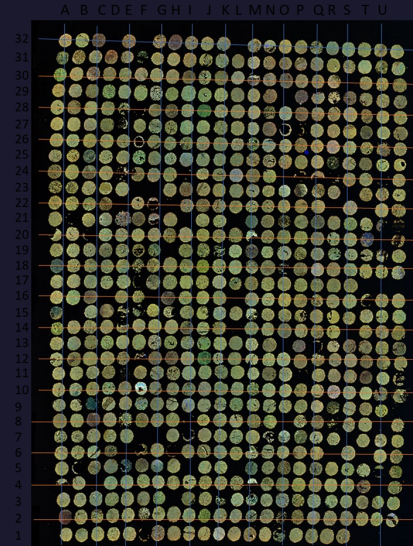
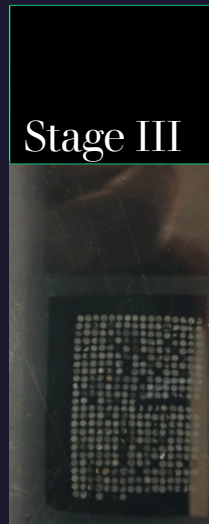
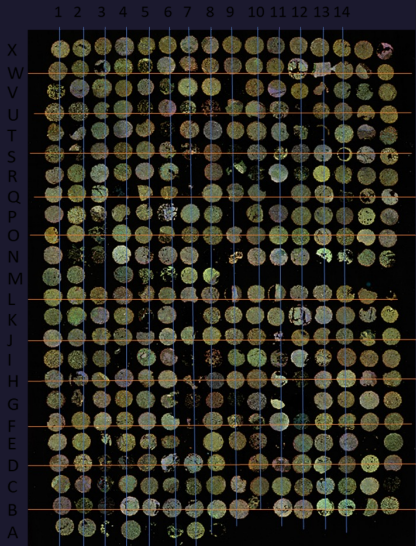


# Current Cohorts to Establish Signatures

Slide	Stage III		Stage IV	
	Fluorescent	Metal	Fluorescent	Metal
Antibody Conjugation	Fluorescent	Metal	Fluorescent	Metal
# of Core (ROI)	384	301	704	640
# of Plex	7	36	7	37
Type of file	.qptiff	.mcd & .txt	.qptiff	.mcd & .txt

IO TMA	
Fluorescent	Metal
77	77
7	36
.qptiff	.mcd & .txt

TIL Samples	
Fluorescent	Metal
130	45
7	36
.qptiff	.mcd & .txt

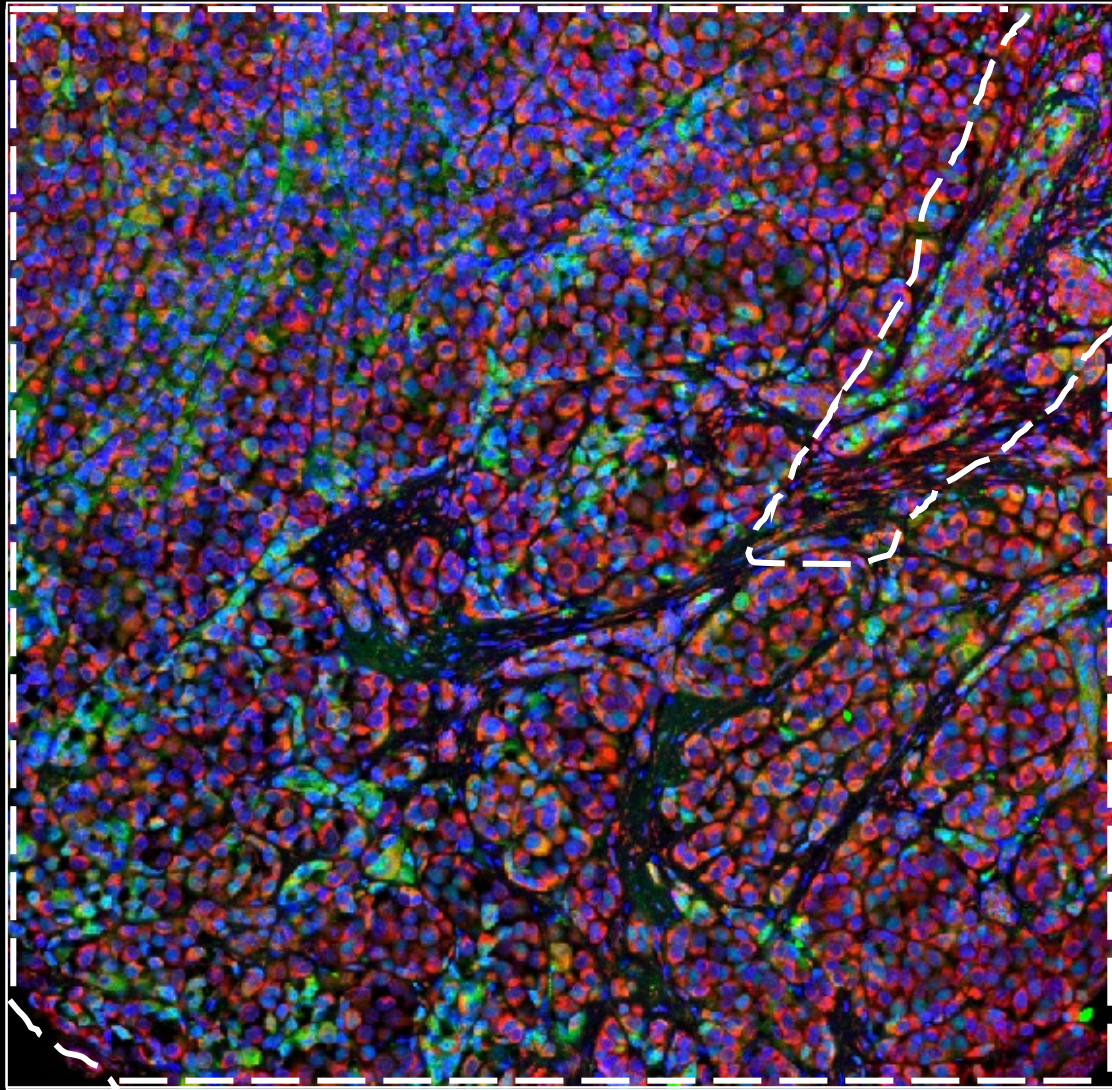


IO TMA 77
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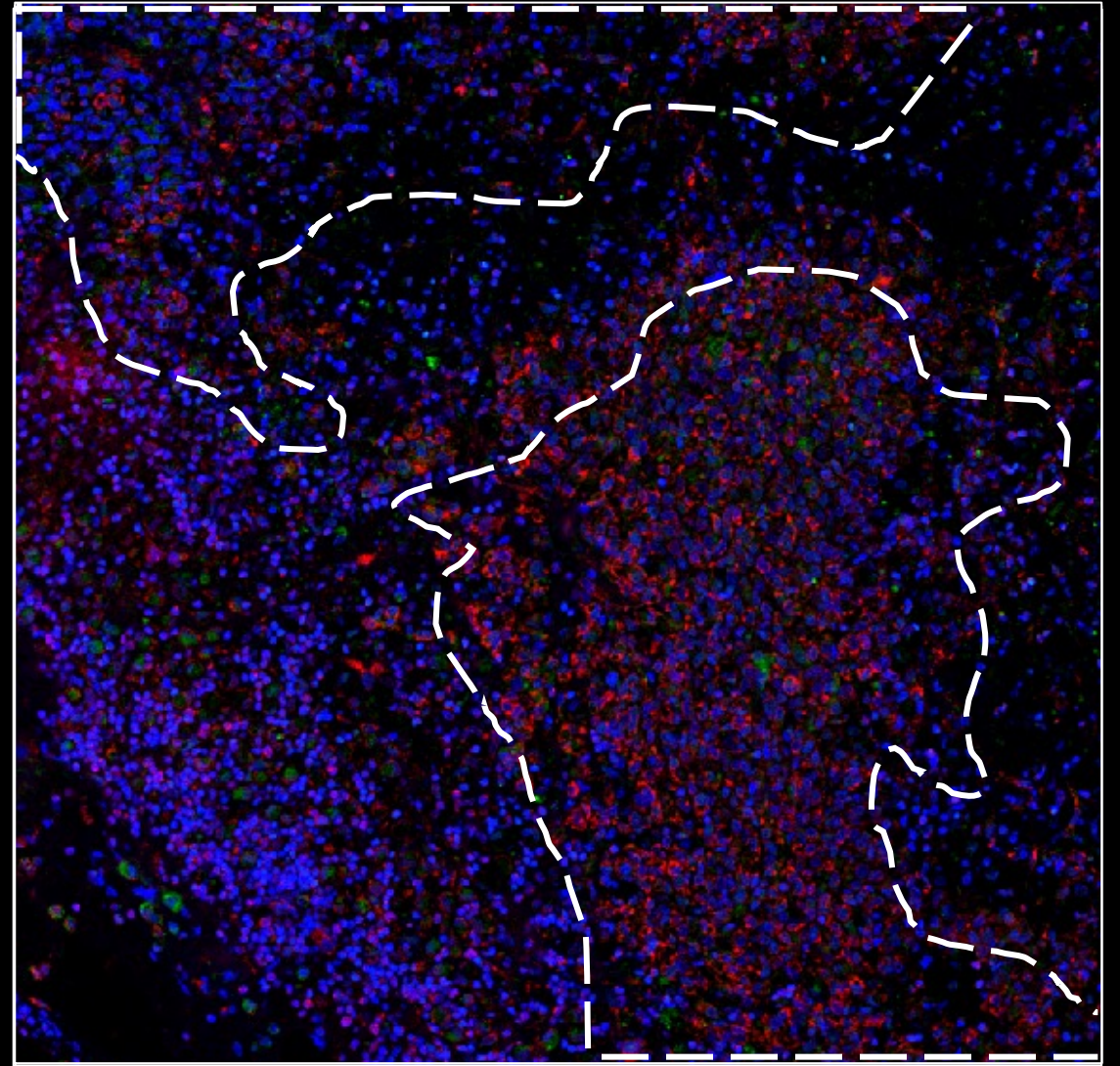
TIL Grow Treated 45	TIL Grow Un Treated 37	TIL No Grow Un Treated 48
------------------------	---------------------------	------------------------------

iNOS; mPGES1

Low TIL



High TIL

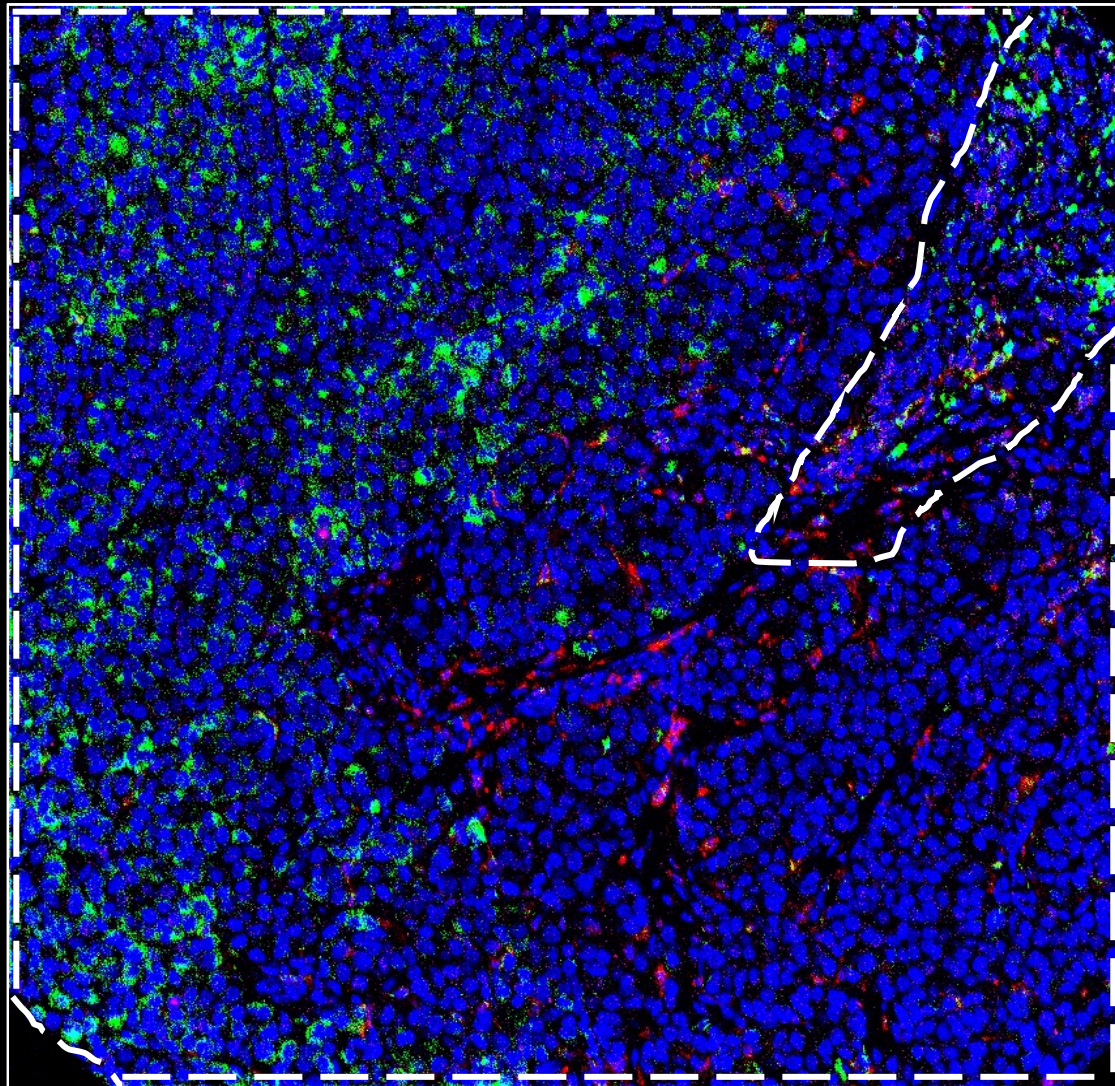


Opal panel

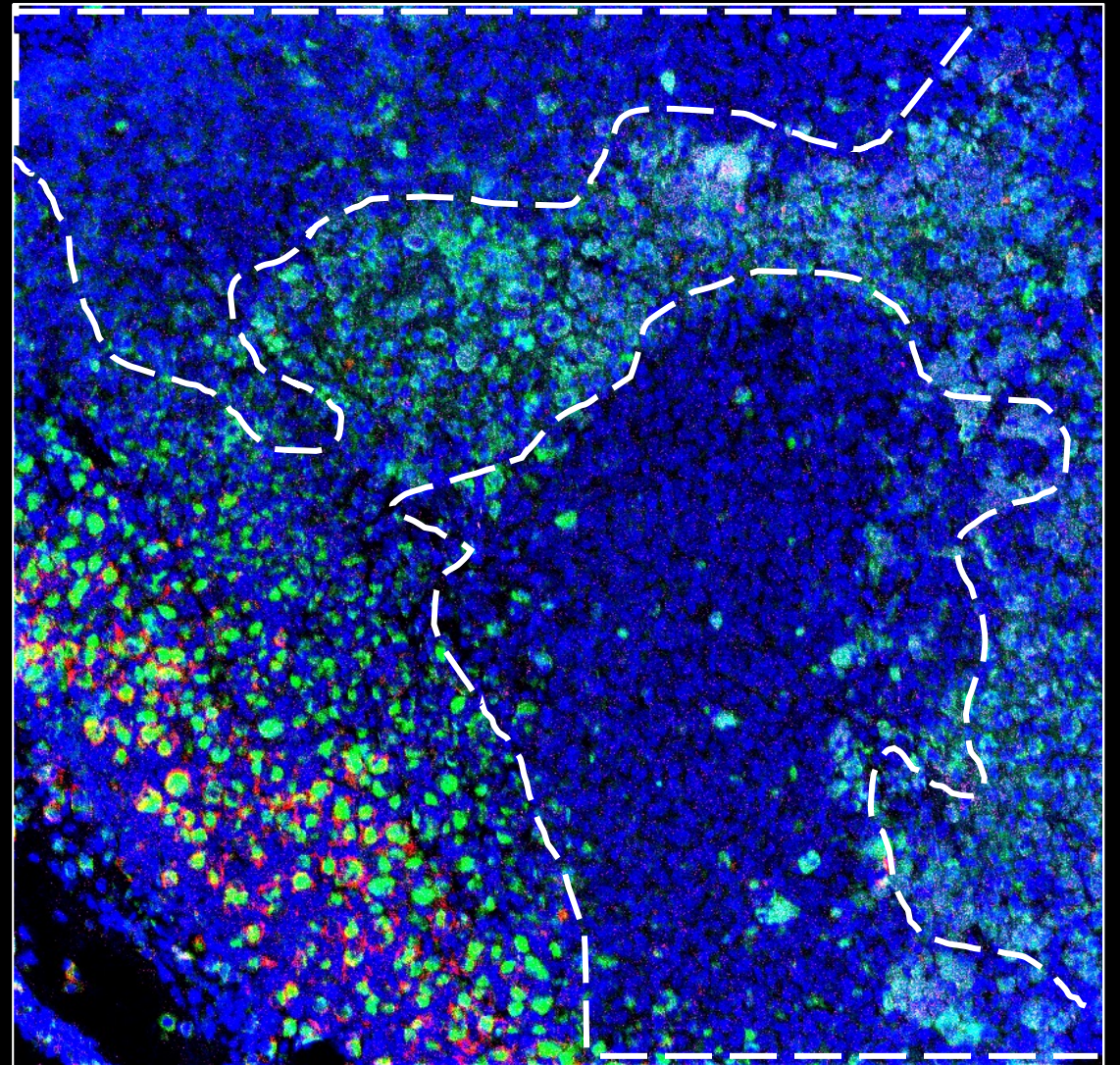
 : Melanoma

CD68; CD163; Arg1

Low TIL



High TIL



IMC panel

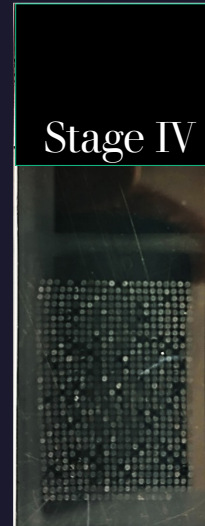
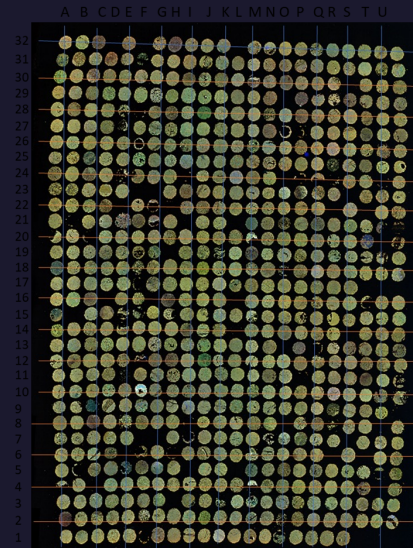
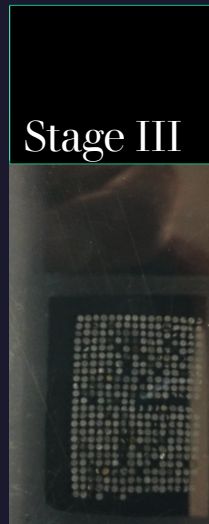
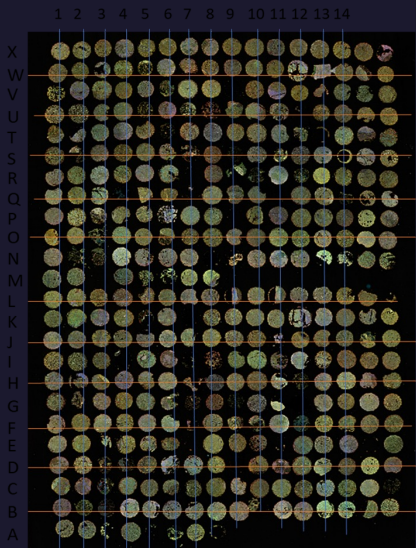
 : Melanoma

# Current Cohorts to Establish Signatures

Slide	Stage III		Stage IV	
	Fluorescent	Metal	Fluorescent	Metal
Antibody Conjugation	Fluorescent	Metal	Fluorescent	Metal
# of Core (ROI)	384	301	704	640
# of Plex	7	36	7	37
Type of file	.qptiff	.mcd & .txt	.qptiff	.mcd & .txt

IO TMA	
Fluorescent	Metal
77	77
7	36
.qptiff	.mcd & .txt

TIL Samples	
Fluorescent	Metal
130	45
7	36
.qptiff	.mcd & .txt

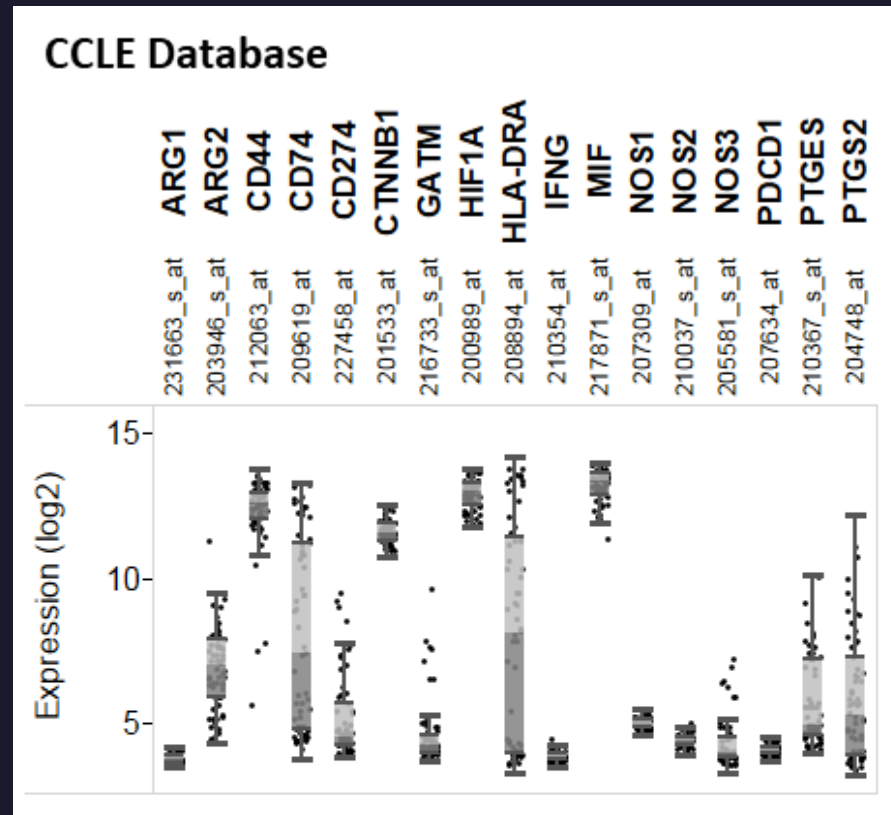


IO TMA 77
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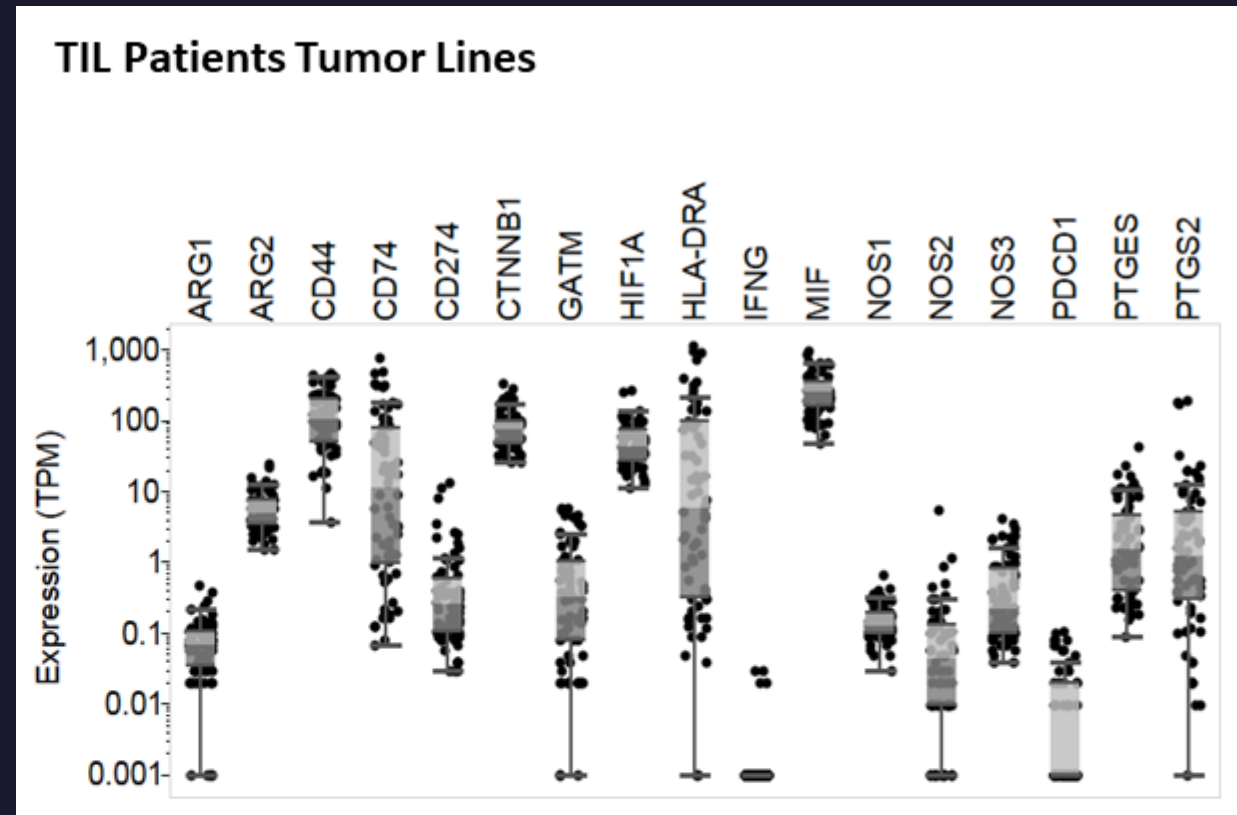
TIL Grow Treated 45	TIL Grow Un Treated 37	TIL No Grow Un Treated 48
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# Initial Findings w/Individual Markers in TIL Treated Patients

## Gene Expression of CD74 Node Markers (CD44, MIF, NOS2, mPGES1) in CCLE and TIL Cohorts



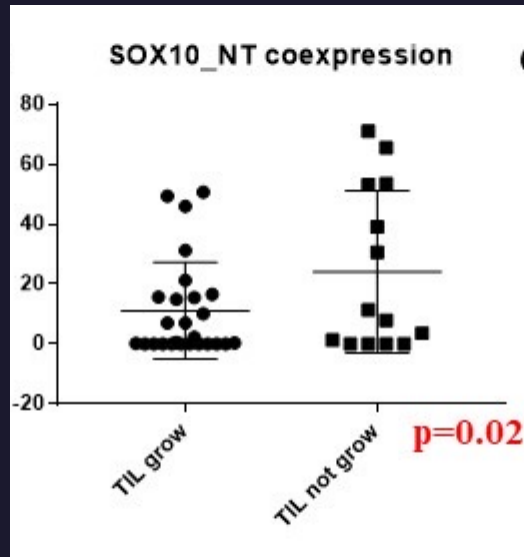
CD74 related genes mRNA expression (based on affymetrix mRNA arrays) in melanoma cell lines from the Cancer Cell Line Encyclopedia (CCLE). The y axis represents the log<sub>2</sub> of the robust multi-array average.



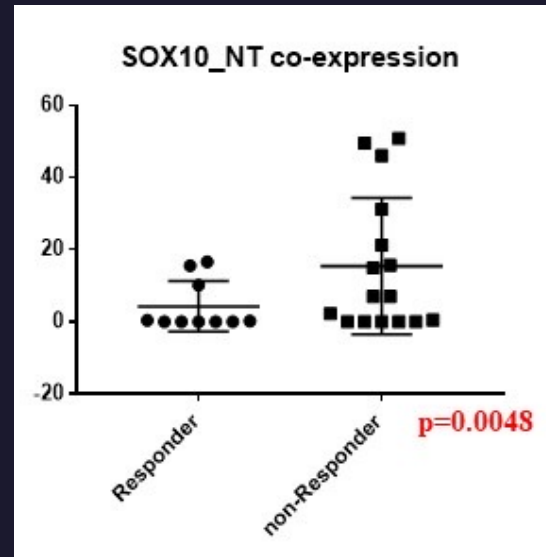
The expression of CD74 related genes mRNA [in transcripts per million (TPM) in MDACC TIL patients' tumor lines dataset.

# Initial Findings w/Individual Markers in TIL Treated Patients

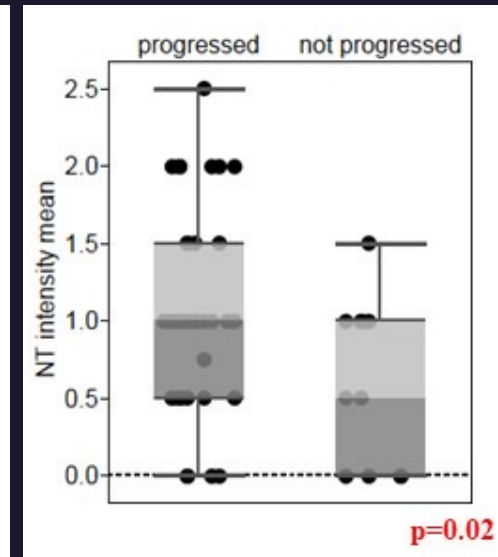
## Tumor Cell NT Expression Correlates with Poor or No TIL Growth from Tumor Samples



**NT expression in tumor cells associated with poor TIL growth.** NT expression in tumor cells in successful TIL growth group, compared to TIL not grow and the significance of this association.



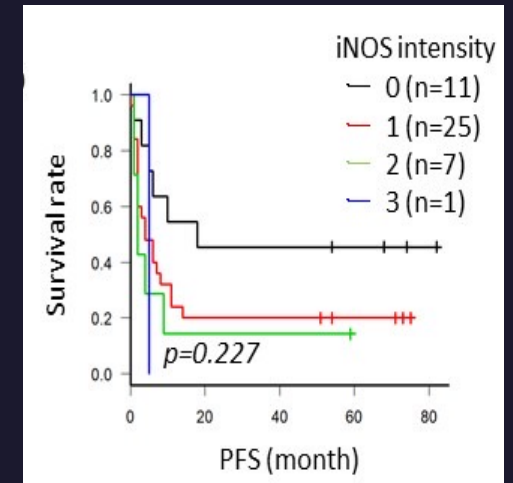
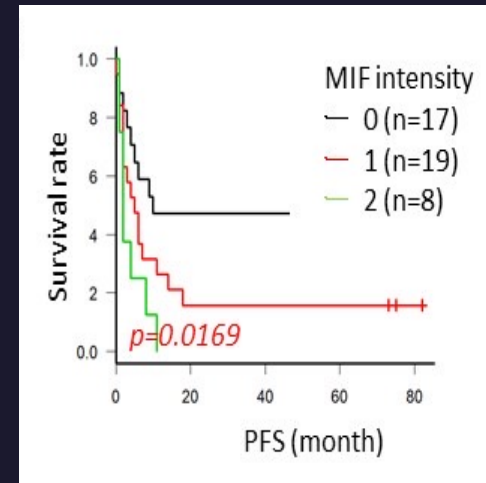
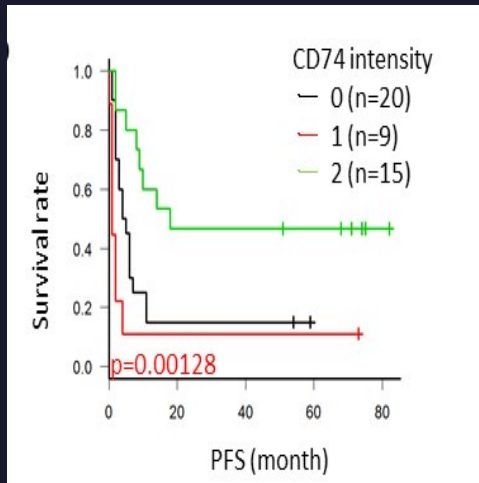
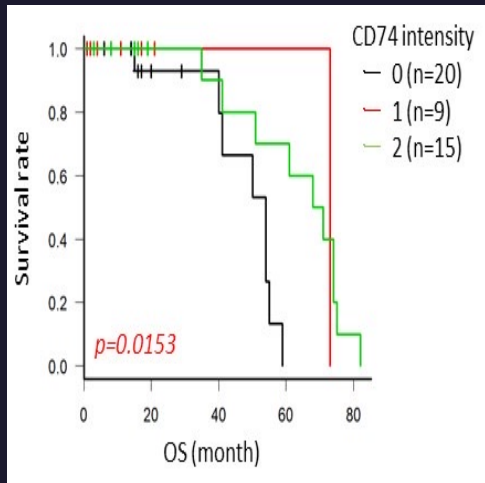
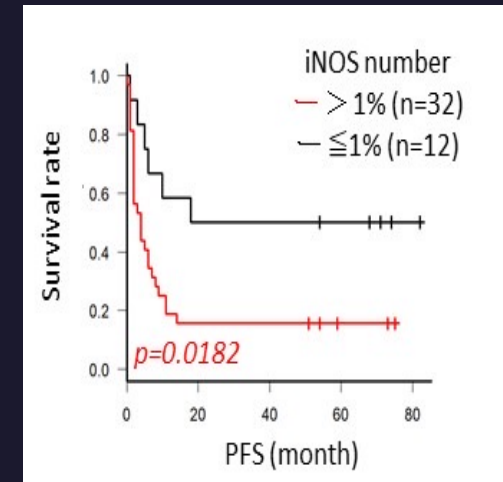
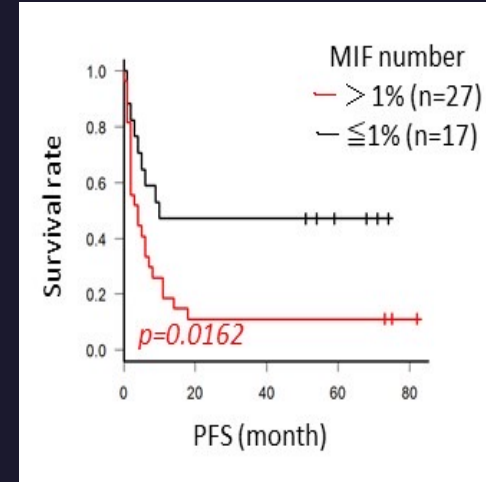
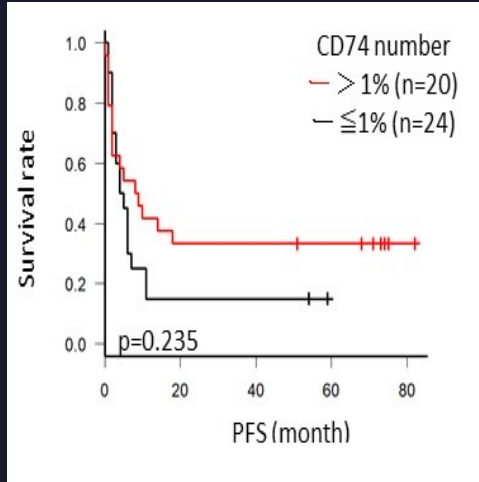
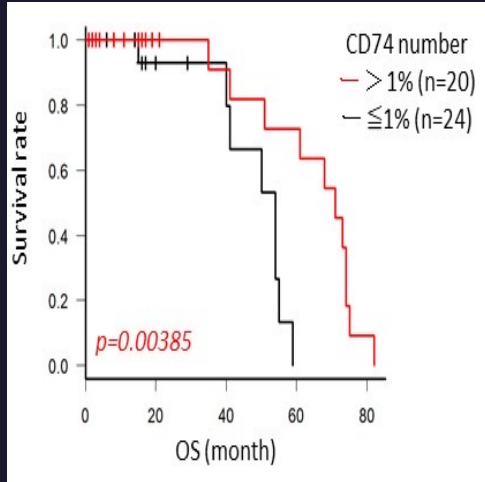
**irRC of TIL Treated Patients and Progression of Disease Associate with NT Expression .** NT expression in tumor cells in responders versus non-responders and their significance of this association. Mean staining intensity of NT expression in progressed patients is significantly high compared to non-progressed patients.



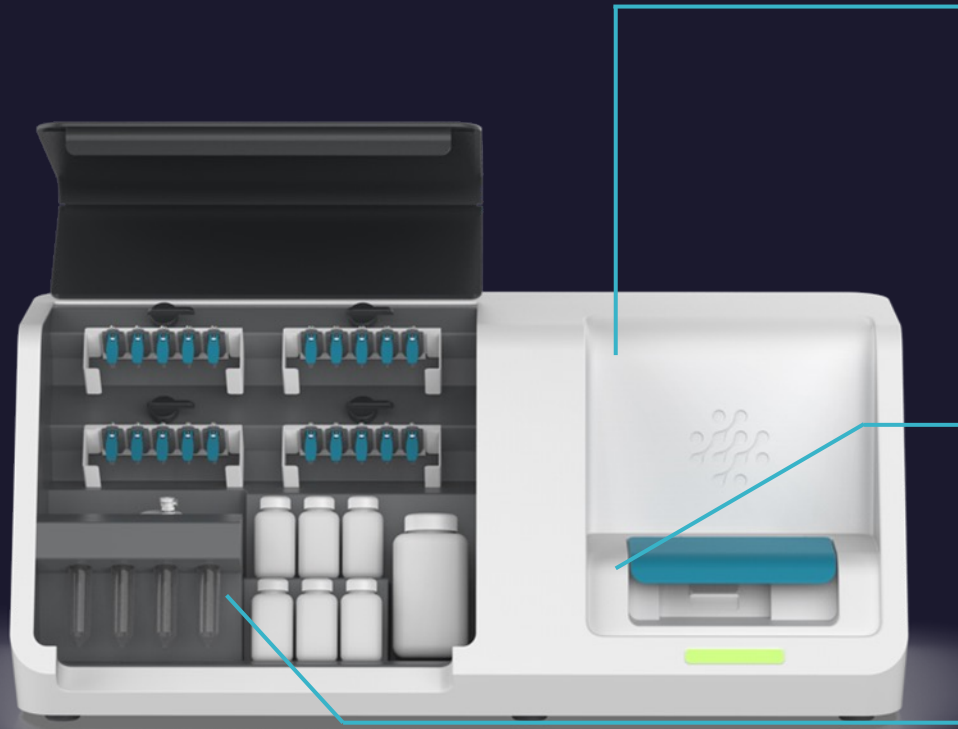
# Initial Findings w/Individual Markers in TIL Treated Patients

Overall and Progression Free Survival by CD74 Number and Intensity

Progression Free Survival by MIF and iNOS Number and Intensity



# Lunaphore Comet



## Microscope

- Fluorescent microscope TRITC, Cy5, DAPI
- 20X - 0.75 NA - 0.23  $\mu\text{m}$  / pixel

## Staining-Imaging Module

- 4 slides tray
- Works with standard histology slides
- Staining-imaging parallelization
- Temperature and pressure control

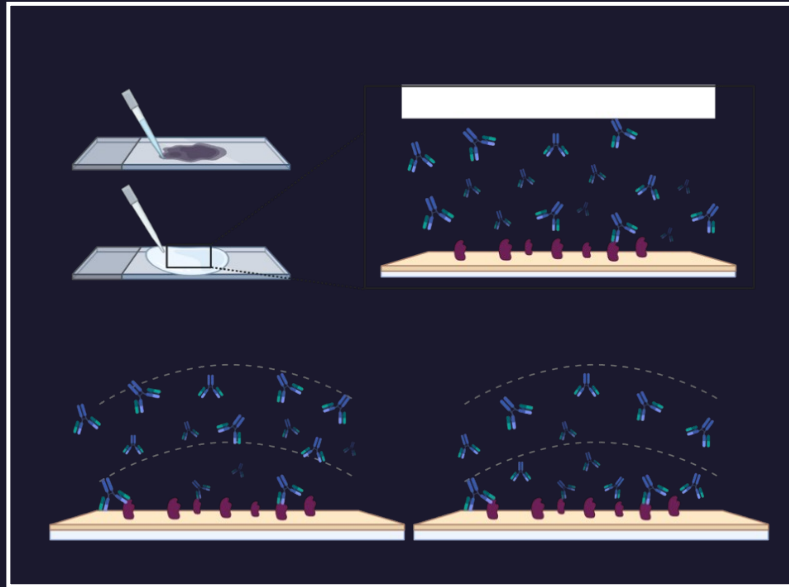
## Reservoirs

- 20 for Ab1; 4 for Ab2; 7 for buffers
- Designed to process 4 slides without refill

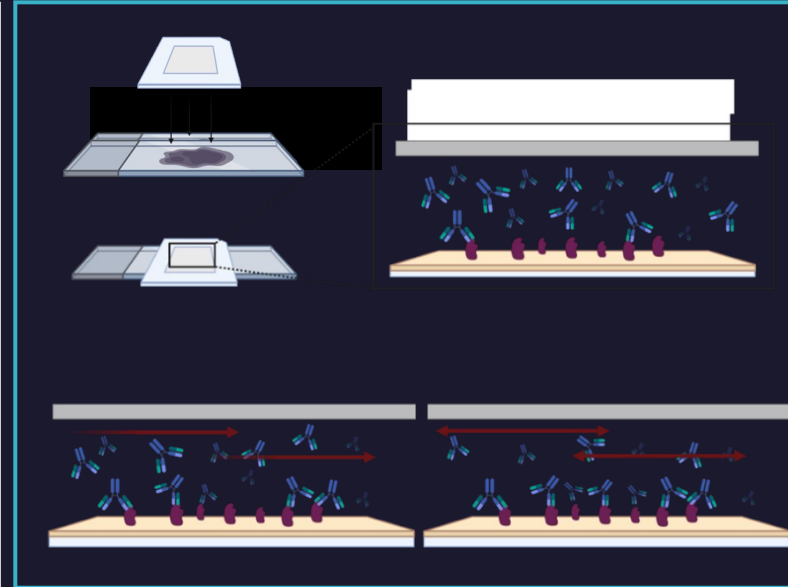


# COMET – Core Chip Technology

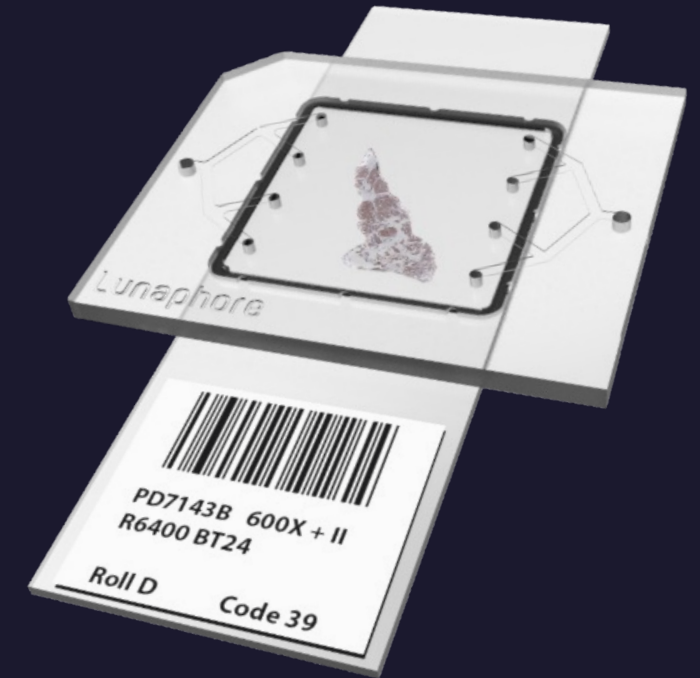
Standard tissue incubation



Fast Fluidic Exchange (FFeX) Technology



- Microfluidic Imaging Chip enables staining & imaging
- Pressure-driven system allowing for fast and uniform delivery of reagents on a tissue section in a closed chamber
- Precisely controlled immuno-reaction within an extremely short incubation time enabled by the FFeX Technology



Slide Modified from Lunaphore

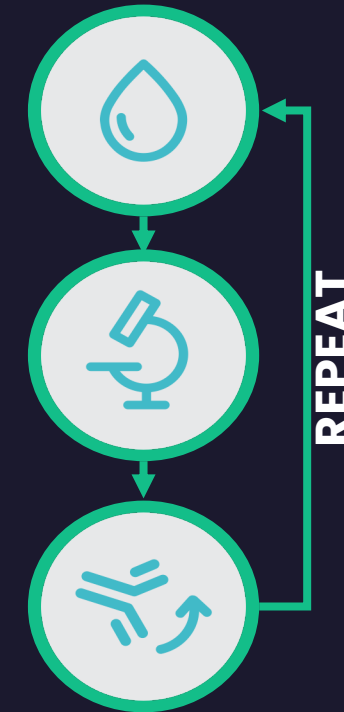
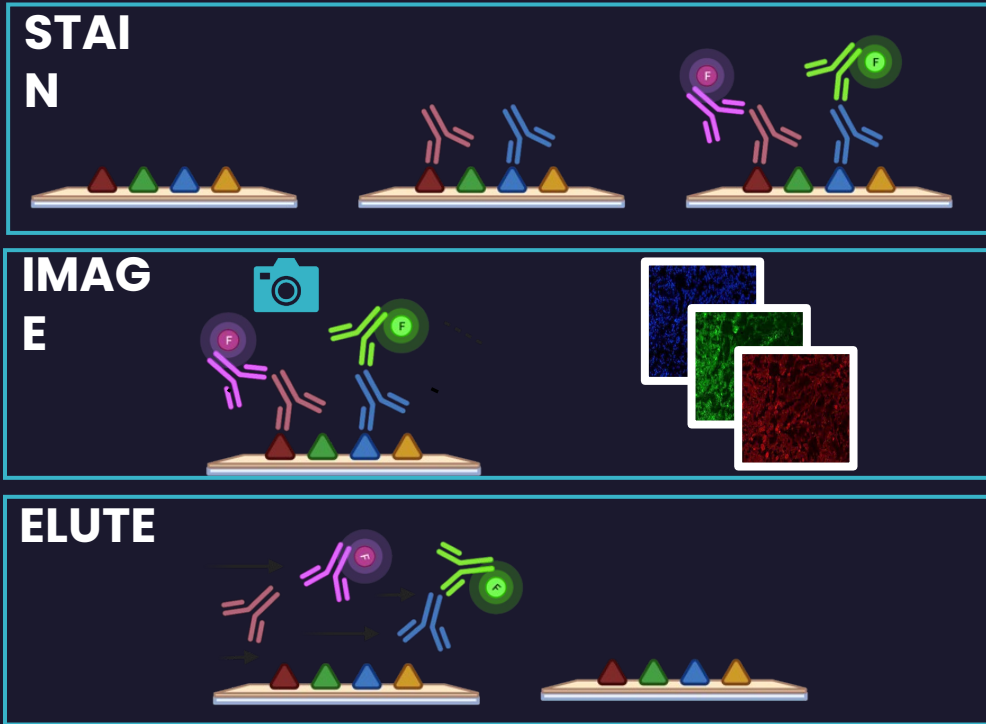
# Protocol Times & Throughput

## PREPARATION

Slide Preparation  
30 min – 2 hours

## AUTOMATED PROTOCOL

Staining (+ Elution) cycle <33 min  
Imaging cycle ~20 min



## Automated Protocol Times

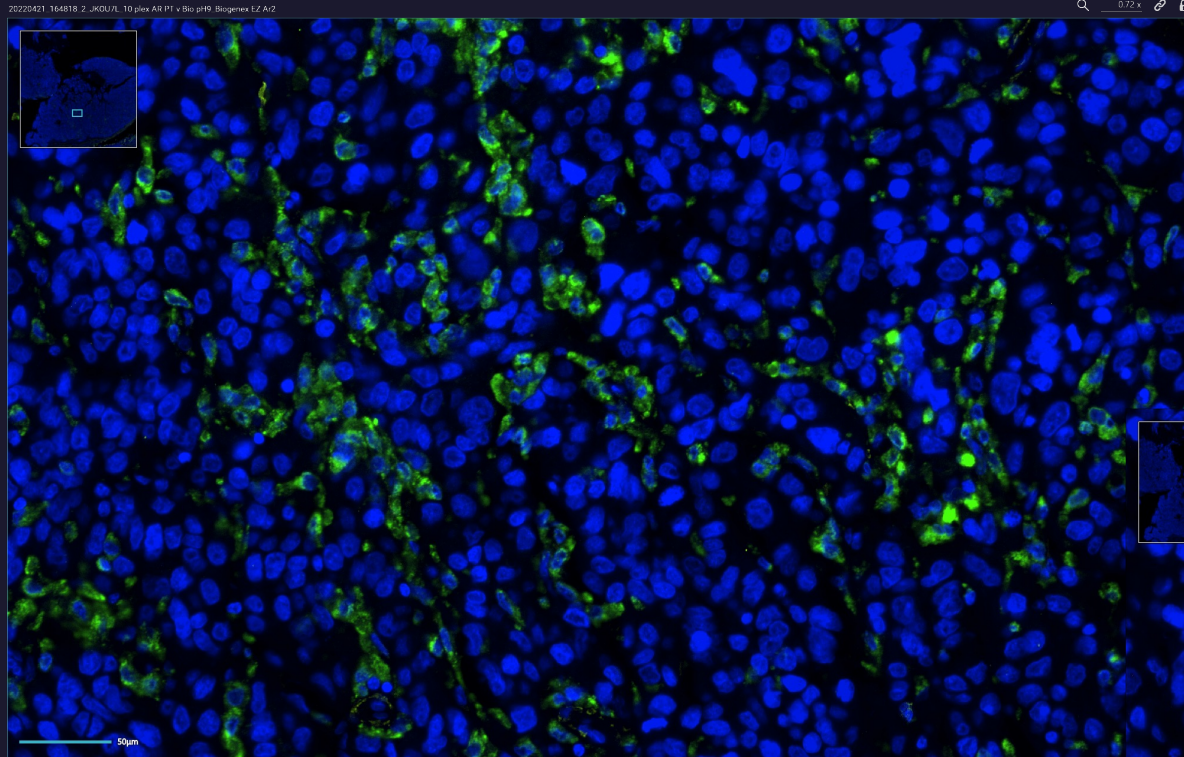
- 10-plex (Antibody Cocktail) on 4 slides: 10-15 hours
- 20-plex (Antibody Cocktail) on 4 slides: 24 hours

## Throughput

- 30 slides / 5 days for 10-plex
- 20 slides / 5 days for 20-plex

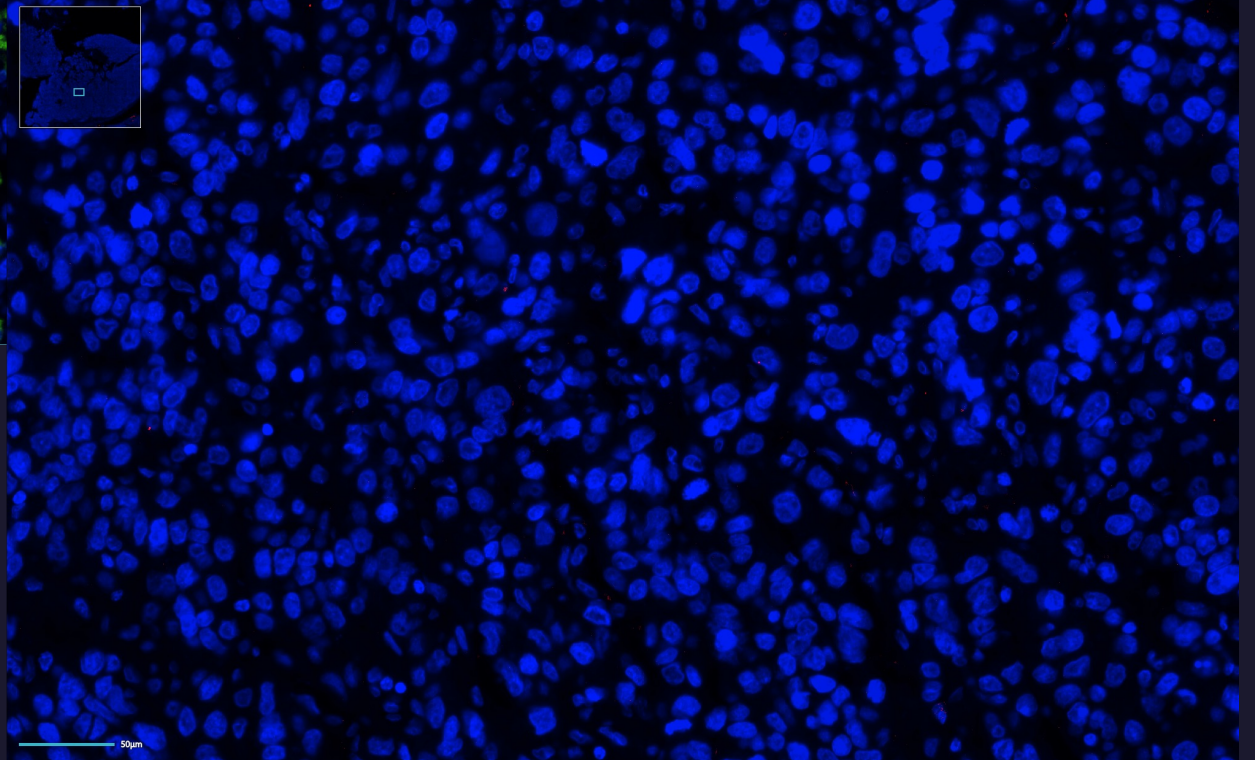
Slide Modified from Lunaphore

# Stain & Elute



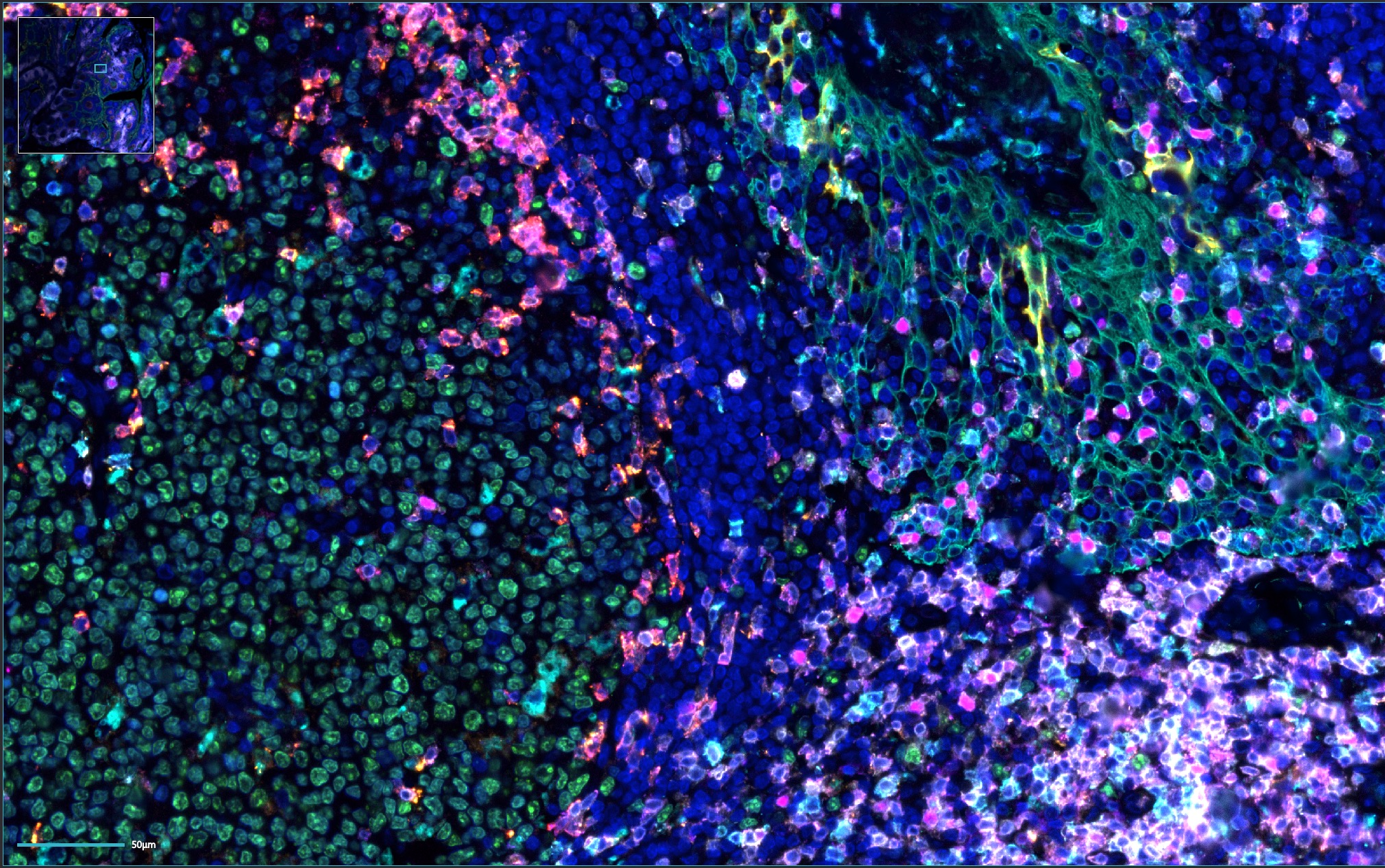
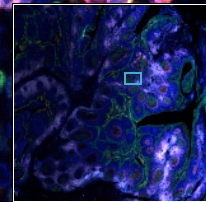
CD68 Stain in Green

CD68 Elution in Red



20220421\_164818\_4\_Mumu16\_Lunaphore 10plex (5 cycles)\_Lunaphore Tonsil

0.72 x



Channels			
	DAPI	DAPI   80 ms	
	Cy5	Cy5   200 ms	
	TRITC	TRITC   400 ms	
	FoxP3	Cy5   200 ms	
	CD68	TRITC   400 ms	
	PD-1	Cy5   200 ms	
	CD8	TRITC   400 ms	
	PD-L1	Cy5   200 ms	
	Ki67	TRITC   400 ms	
	CD4	Cy5   200 ms	
	CK	TRITC   400 ms	
	CD3	Cy5   200 ms	
	CD20	TRITC   400 ms	

Annotations  
Info



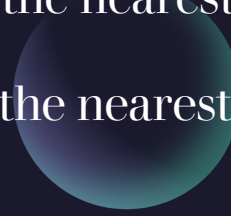
# Significant Findings

Our proposed markers of oxidative stress and immune-related enzymes and their mediators defines TIME architecture and predicts overall survival of advanced stage melanomas.

- Most of the melanoma cells reside in iNOS/mPGES1 NBH in both LTS and STS in Stage III and IV melanomas
- iNOS/mPGES1 and iNOS/NT expressing cells reside significantly closer to each other than additional inflammatory profiles.

Inflammatory signatures (defined by iNOS/NT/mPGES1, and CD74/CD44/MIF expression characteristics) regulates immune profiles;

- There is a significantly higher average proportion of CTLs in LTS than in STS
- Anti-tumor immune cells (NK and CTLs) are more clustered together in LTS than STS.
- The average distance from a tumor community to the nearest CTL community is shorter in LTS than STS.
- The average distance from an average M2 TAM to the nearest B cells is longer in LTS than STS in Stage IV melanomas
- The proportion of M2 TAM cells that are also in the MIF CD44 NBH, out of all M2 TAM cells is higher in STS than in LTS



# Future

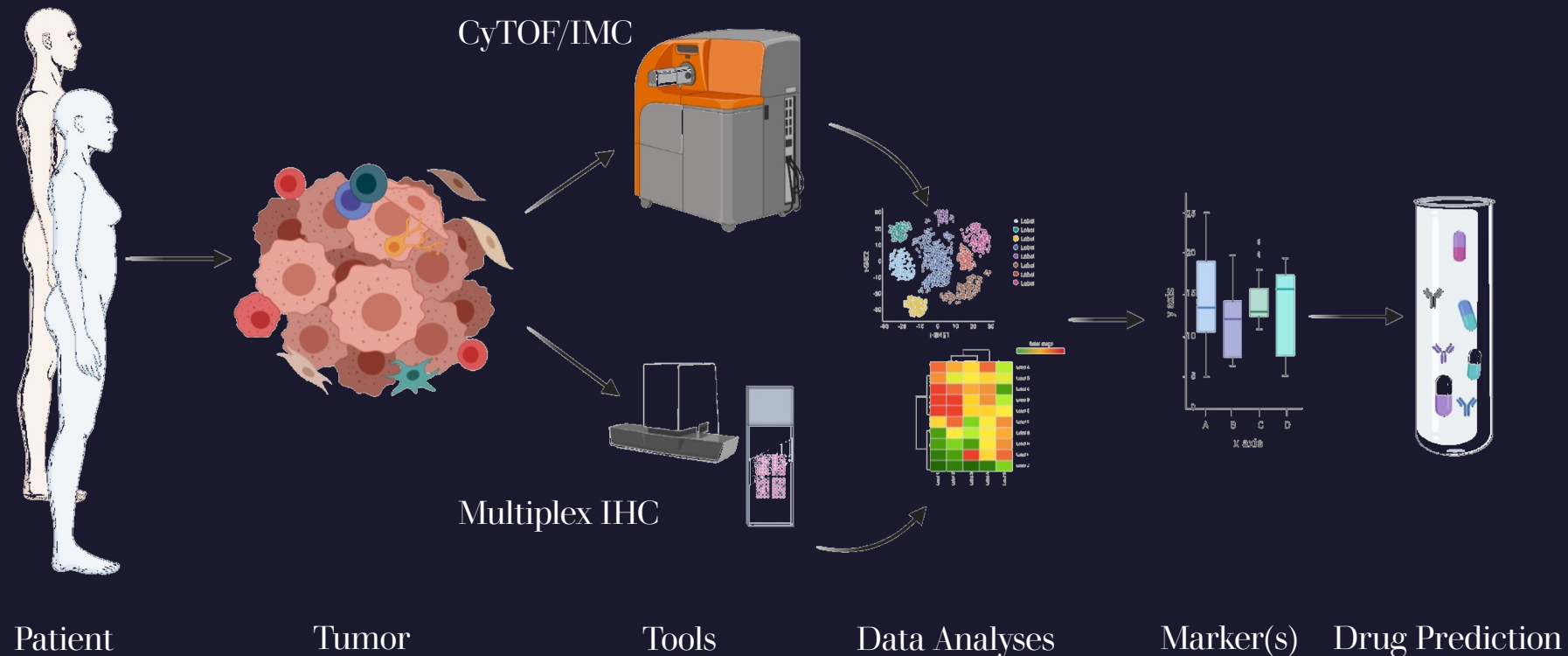
More to Do – *As always...*

Correlate TIME signatures with multiscale radiomic properties that can be derived from routine CT scans to inform a mathematical model for the early prediction of the response to IO agents.

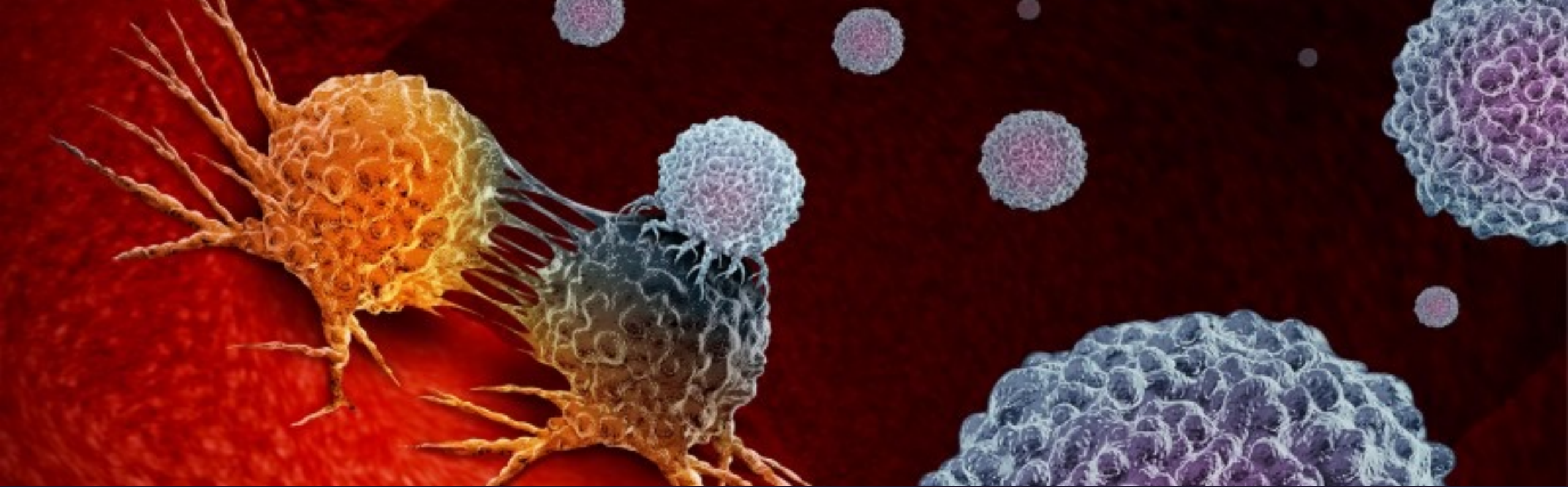
Extend and validate the mathematical model to predict melanoma response to immunology (IO) agents using the following data types;

- Imaging data: standard of care imaging, including pre-immunotherapy (T0) and post-immunotherapy (T1, T2, etc.) CT scans to quantify lesion volume and volume change over time. Our typical approach in the clinic is to obtain CT scans once every 3 months.
- Multiplex IHC data: We will correlate the refined set of markers in the mIF and CyTOF immune signatures, including iNOS, CXCR4, and CXCR7 for tumor cell proliferation rates (model parameter  $\alpha$ ), IFN $\gamma$ R1, CD3+, CD4+, CD8+ and macrophage markers CD11b+F4/80+CD11c-Ly6G- as quantitative measures of tumor immune infiltration (model parameter  $\Lambda$ ); and CD44 and dendritic cell CD11c+Ly6G-F4/80-, and MDSC markers CD11b+Gr-1Ly6G+ as quantitative indicators of immune cell kill efficacy (model parameter  $\mu$ ).

Validate each marker individually, and then in sets signatures, for prognostic and then by testing of predicting response to immunotherapy using (ongoing as well as retrospectively collected) human melanoma (and others) biopsy samples from patients with known immunotherapy outcomes.







# Summary

Our platform serves as comprehensive “molecular diagnosis” tool to support precision medicine approach by providing access to high-density proteomics and radiomics information.



# Team

**Sungnam Cho, MS**  
**Yasunari Fukuda, MD, PhD**  
**Dai Ogata, MD, PhD**  
**Sun-Hee Kim, PhD**  
**Joel Eliason, GradStudent**



MDACC Flow Cytometry and  
Cellular Imaging Core Facility  
**Jared K. Burks, PhD**

MDACC MelCore  
**Lauren Haydu, PhD**  
Jared Malke  
Sheila Duncan

MDACC Pathology Team  
**Carlos Torres Cabala, MD**  
Victor G. Prieto, MD, PhD

Melanoma Medical Oncology  
Elizabeth A. Grimm, PhD

Saint John's Cancer Institute  
**Matias A. Bustos, PhD**  
Dave S.B. Hoon, PhD



**Mike Spencer, Ph.D.**  
*Director of IHC & Digital Path*

Computational Medicine and Bioinformatics Team  
University of Michigan  
**Arvind Rao, PhD**

Radiomics Project Team

**Eugene J Koay, MD., PhD** – MDACC, Radiation Oncology  
Vittorio Cristini, PhD - Methodist Research Institute, Mathematical Medicine  
Adi Diab, MD – MDACC, Melanoma Medical Oncology

MDACC Investigational Cancer Therapeutics  
**Aung Naing, MD, FACP**  
Funda Meric-Bernstam, MD



**T. Regan Baird, PhD**  
**Dan Winkowski, PhD**

# Thank You

Suhendan Ekmekcioglu, PhD

[sekmekcioglu@mdanderson.org](mailto:sekmekcioglu@mdanderson.org)

